

Scientific Transactions in Environment and Technovation

# In vitro biological control of Fusarium oxysporum f. sp. cubense (Foc), casual agent of wilt disease in Musa paradisiaca L.

## S. Thevasundari

Department of Botany, Seethalakshmi Ramaswami College, Tiruchirappalli - 620 002.

## Abstract

Fusarium wilt of banana, also called Panama disease, is one of the most important and destructive diseases of banana plant (*Musa paradisiaca* L.) in the world, which has been reported from all banana producing countries. Control of *Fusarium oxysporum* f. sp. *Cubense*, the casual agent of wilt disease, by using the plant extracts has been evaluated as a strategy of biological control. The antifungal activity of different extracts of *Justicia adhatoda* L., *Ocimum tenuiflorum* L., *Piper betle* L. and *Vitex negundo* L. against *F. oxysporum* f. sp. *cubense* was evaluated by well diffusion method. The results revealed that the plant extracts of *V. negundo* L. exhibited maximum antifungal activity. *In vitro* screening of plant extracts showed encouraging results, and hence they could be potentially used in the management of *Fusarium* wilt in banana plant caused by *F. oxysporum* f. sp. *cubense*.

Key words : Fusarium wilt, Musa paradisiaca L., biological control, plant extracts

Received : July 2015

## INTRODUCTION

Banana (*Musa* sp.) is one of the first cultivated food crops. It is the fourth most important global food commodity after rice, wheat and maize in terms of gross value production. At present, it is grown in more than 150 countries throughout the tropical and subtropical regions, and it is the staple food for more than 400 million people (Mohapatra *et al.*, 2010). In India, main banana growing states are Tamil Nadu, Maharashtra, Karnataka, Gujarat, Andhra Pradesh, Bihar, Assam and Madhya Pradesh. Tamil Nadu is one of the leading banana producing states in India which contributes 19% of the total production (Indian Horticulture database, 2014).

Fusarium wilt of banana, also called Panama disease, is caused by *Fusarium oxysporum* f. sp. *cubense* (E.F. Smith) Snyder and Hansen, shortly as Foc (Ploetz, 2006). Fusarium wilt is known to be one of the most important and destructive diseases of banana. It has been reported in all banana-producing countries, including Asia, America, Africa, and Australia (Ploetz and Churchill, 2011) and it was responsible for losses of billions of dollars in the first export trades in the world (Jones, 2009; Ploetz *et al.*, 2015).

The use of botanicals has a long history in the control of plant diseases. Plant biochemicals and crude extracts have been reported to have antimicrobial properties against plant pathogens including fungi,

\*Corresponding Author : email: thevasundari@gmail.com bacteria, nematodes and insects *in vitro* as well *in vivo*. Further, the use of botanicals is regarded as the best suited environmental friendly measure as they are easily biodegradable and safer (Pradhanang *et al.*, 2003). Consequently the present investigation was designed to assess the *in vitro* biological control of *F. oxysporum* f. sp. *cubense* casual agent of wilt disease in *Musa paradisiaca* L. by using some medicinal plants.

Revised and Accepted : July 2017

## MATERIALS AND METHODS

## Collection of Banana Plant

The Fusarium wilt disease affected banana pseudo stem (Banana variety - Poovan) was collected from banana cultivated field of Marungulam, Thanjavur district.

## Isolation of plant pathogen

The pathogen *F. oxysporum* f. sp *cubense* (FOC), was isolated from Fusarium wilt disease affected banana pseudo stem (*M. paradisiaca* L.) by using potato dextrose agar (Himedia, Mumbai) medium. The identification of pathogen was done by macro and micro morphological examinations by using the standard manuals - A Manual of Soil Fungi (Gillman, 1957) and Fungi in Agricultural Soils (Domsch and Gams, 1972).

## **Collection of plants**

Fresh plant leaves of *Justicia adhatoda* L., *Ocimum tenuiflorum* L., *Piper betle* L. and *Vitex negundo* L. were collected from the natural habitat of Marungulam,

July to September 2017

P - ISSN 0973 - 9157 E - ISSN 2393 - 9249

#### J. Sci. Trans. Environ. Technov. 11(1), 2017

Thanjavur district, South India. The plant species was identified and confirmed with reference to Herbarium specimens available in the Rapinat Herbarium, St. Joseph's College, Tiruchirappalli, Tamil Nadu, India. These plants were selected based on traditional knowledge.

#### **Preparation of plant extracts**

Plant extracts were prepared following the methodology of Indian Pharmacopoeia (Indian Pharmacopoeia, 1996). The plant leaves were shade dried and pulverized. Then, 50g of each leaf powder was packed separately in Soxhlet apparatus and subjected to continuous hot percolation for 8 h using 450 mL of ethyl acetate, methanol, n-hexane and petroleum ether as solvents. The obtained extracts were concentrated by using a rotary flash evaporator. The plant extracts were preserved in airtight containers for further analysis.

#### Screening of antifungal activity

The antifungal activity of the selected medicinal plants was tested against F. oxysporum f. sp cubense by using agar well diffusion method (Perez et al., 1990). The freshly prepared and sterilized potato dextrose agar medium was supplemented with streptomycin (10 µg/ mL) to prevent bacterial contamination, poured into each Petri plate and allowed to solidify. The test fungal culture was evenly spread over the media by using sterile cotton swab. Then wells (5 mm) were made in the medium by using sterile cork borer, 200 µl of ethyl acetate, methanol, n-hexane and petroleum ether extracts of selected medicinal plants were transferred to wells in separate plates. The commercial fungicide carbendazim (mg/ml) and solvents (ethyl acetate, methanol, n-hexane and petroleum ether) were used as positive and negative controls respectively. Then these plates were incubated at 28 °C for 48-72 h. After incubation period the results were observed and the diameter of inhibition zone around each well was measured. All the tests were performed under aseptic condition, three replicates were maintained and the activity was expressed as the mean of inhibition diameters (mm) produced by the plant extracts.

## **RESULTS AND DISCUSSION**

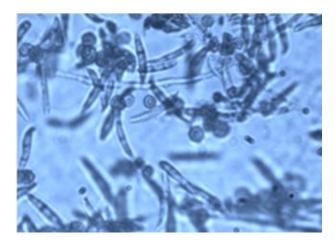
In the present study, the plant pathogen, *F. oxysporum* f. sp *cubense*, was isolated from infected stem of banana plant using PDA medium. The pathogen macroscopically appeared as white myulial mat over the medium and microscopic structures revealed the presence of macro conidia and micro conidia (Fig. 1 and 2).

P - ISSN 0973 - 9157 E - ISSN 2393 - 9249

July to September 2017



Figure 1. Macroscopic view of *F. oxysporum* f. sp. *cubense* 



## Figure 2. Microscopic view of F. oxysporum f. sp. cubense

In the present day constraints of chemical pesticides on the environment and human health attempts have been made to find alternative methods to manage the plant diseases and pathogens, the biological methods including use of antagonists, soil augmentation with organic residues, green manures, etc. have been recognized as viable alternatives.

The plant world is a rich store house of natural chemicals that could be exploited for the use as fungicides. The presence of antifungal compounds in higher plants has long been recognized as important factors for controlling some plant diseases (Tapwal *et al.*, 2011). Plant bio-fungicides are cheap, locally available, non toxic and are easily degradable. There is a great demand for them as alternative agents to control plant pathogenic fungi (Singh and Srivastava, 2013).

#### 44 S. Thevasundari

In the present investigation, four plants such as J. adhatoda L., O. tenuiflorum L., P. betle L. and V. negundo L. were screened in vitro for antifungal activity against phytopathogenic fungus, F. oxysporum f. sp cubense. The results of antifungal efficacy of ethyl acetate, methanol, n-hexane and petroleum ether extracts of J. adhatoda L., O. tenuiflorum L., P. betle L. and V. negundo L. are presented in Table 1. Among the four plants tested, the plant extracts V. negundo L. exhibited maximum antifungal activity against *F. oxysporum* f. sp cubense followed by J. adhatoda L. and P. betle L. extracts. O. tenuiflorum L. plant extracts exhibited least antifungal activity against the tested plant pathogen. The ethyl acetate extract of all the plants showed the maximum antifungal activity and petroleum ether extract showed minimum activity against the test pathogen. The results of antifungal effect of four solvents such as ethyl acetate, methanol, n-hexane and petroleum ether revealed no activity against test pathogen. The commercial fungicide carbendazim showed the highest antifungal activity against F. oxysporum f. sp cubense (23 mm). The antifungal activity of ethyl acetate extract of V. negundo L. was found to be comparable to the standard fungicide tested.

In the present study, *V. negundo* L. plant extracts exhibited highest antifungal activity against *F. oxysporum* f. sp *cubense*. The results of antifungal activity of *V. negundo* L. were in harmony with results of Siva *et al.* (2008) who reported that *V. negundo* L. plant extracts exhibited significant antifungal activity against *F. oxysporum* causing wilt disease of *Solanum melogena* L. in *in vitro*. Recently a study by Bapat *et al.* (2016) reported considerable antifungal activity of ethanolic and petroleum ether extracts of *V. negundo* L. against phytopathogenic fungus, *Sclerotium rolfsii* Sacc.

In the same way, antifungal activity of various medicinal plants against *F. oxysporum* has been reported by several researchers. Dwivedi and Enespa (2012) reported that *Moringa oleifera* extracts (leaves, bark and seeds) showed significant inhibition in the mycelial growth of *F. oxysporum* f. sp. *lycopersici*. In addition, Huang *et al.* (2012) reported the inhibitory effects of *Allium tuberosum* on *F. oxysporum* f. sp. *cubense*. Hadi *et al.* (2013) also accounted *Mentha piperita* plant extracts exhibited remarkable antifungal activity against *F. oxysporum*. Recently, Neela *et al.* (2014) who reported that *P. betle* L. plant extracts exhibited significant antifungal activity against *F. oxysporum* f. sp. *lycopersici* the causal agent of Fusarium wilt in tomato.

In the present study it was also found that the plant extracts of *J. adhatoda* L. showed moderate antifungal activity against *F. oxysporum* f. sp. *cubense.* This is in

P - ISSN 0973 - 9157 E - ISSN 2393 - 9249

July to September 2017

contrast to the finding of Siva *et al.* (2008) who reported strong fungicidal activity of *Adhatoda vasica* against *Fusarium oxysporum* causing wilt disease in *Solanum melogena* L.

The extracts of *O. tenuiflorum* L. were found to have least antifungal activity against *F. oxysporum* f. sp. *cubense*. On the contrary, mycelial growth of various plant pathogens such as *Botryodiplodia theobromae* causing fruit rot of mango (Patil *et al.*, 1995), *B. theobromae* causing fruit rot of sweet orange (Godara and Pathak, 1995), *F. oxysporum* causing wilt disease of *Solanum melogena* L. (Siva *et al.*, 2008), *F. solani* causing dry rot disease of potato (Bhardwaj, 2012) and *Alternaria solani* which causes early blight disease on tomato plants (Dheeba *et al.*, 2015) was inhibited by *O. tenuiflorum* L. plant extracts. This could be due to the fact that there is variations in the sensitivity of micro organisms to different plant extracts.

The results indicated that all the tested plant extracts had different levels of antifungal activity against the phytopathogenic fungus *F. oxysporum* f. sp. *cubense*. The differences in the inhibitory effect of tested plant extracts may be due to qualitative and quantitative differences in the antifungal principles or compounds present in them.

## Table 1: *In vitro* biological control of *F. oxysporum* f. sp *cubense* by using some medicinal plants

		Zone of inhibition (Diameter in mm)			
SI.	Name of the	Ethyl	Methanol	n-Hexane	Petroleum
No.	Plant	acetate	extract	extract	ether
		extract		CALLOC	extract
1	J. adhatoda L.	15.7 ± 2.52	14 ± 1	12.3 ± 0.58	11±1
2	O. tenuiflorum L.	11.3 ± 1.12	11±1	10±1	9.3 ± 0.6
3	Piper betle L	14.3 ±1.15	12.3±1.53	11.3±0.58	10±1
4	V. negundo L.	29±0.5	17± 0.82	14.7 ± 1.53	12±1

Results expressed as Mean ± Standard Deviation (n-3)

## CONCLUSION

The findings of this study suggest that *V. negundo* L. plant extracts possessed broad spectrum antifungal activity against *F. oxysporum* f. sp. *cubense*. This potential could be exploited for the an ecofriendly management and control of wilt disease of banana plant. In addition, *V. negundo* L. plant extracts could be applied as agro-eco-friendly fungicides for various crops.

## REFERENCES

Bapat U.C., Smrity Prabha and Jyoti Kumar, 2016. Antifungal activity of ethanolic and petroleum ether extracts of some medicinal plants against the plant pathogenic fungus *Sclerotium rolfsii* sacc. *Int. J. Bioassays*, 5(7): 4714-4719.

#### https://doi.org/10.21746/ijbio.2016.07.0012

- Bhardwaj, S. K. 2012. Evaluation of plant extracts as antifungal agents against Fusarium solani (Mart.) Sacc. World J. Agric. Sci., 8 (4): 385-388.
- Dheeba, B., Niranjana, R., Sampathkumar, P., Kannan, K. and Kannan, M. 2015. Efficacy of Neem (*Azadirachta Indica*) and Tulsi (*Ocimum Sanctum*) leaf extracts against early blight of tomato. *Proc. Natl. Acad. Sci.*, *India, Sect. B Biol. Sci.*, 85: 327. https://doi.org/10.1007/s40011-014-0340-9
- Domsch, K.H. and Gams, W. 1972. Fungi in agricultural soils. Halsted Press Division, New York, USA.
- Dwivedi, S.K. and Enespa, A. 2012. Effectiveness of extract of some medical plants against soil borne fusaria causing diseases on Lycopersicon esculantum and Solanum melongena. Int. J. Pharm. Bio. Sci., 3(4):1171-1180.
- Gillman, J.C. 1957. A Manual of Soil Fungi. Revised 2nd edition Oxford and IBH publishing company (Indian reprint) Calcutta, Bombay, New Delhi.
- Godara, S.L. and Pathak, V.N. 1995. Effect of plant extracts on post harvest rotting of sweet orange fruits. *Indian J. Mycol. Plant Pathol.*, 25(2): 134-135.
- Hadi, M., Kashefi, B., Sobhanipur, A. and Rezaarabsorkhi, M. 2013. Study on effect of some medicinal plant extracts on growth and spore germination of *Fusarium* oxysporum schlecht. in vitro. American Eurasian J Agric and Environ Sci., 13(4): 581-588.
- Huang, Y. H., Wang, R. C., Li, C. H., Zuo, C. W., Wei, Y. R., Zhang, L. and Yi, G. J. 2012. Control of Fusarium wilt in banana with Chinese leek. *Eur. J. Plant Pathol.*, 134(1): 87–95. PMid:23144534 PMCid:PMC3491907 <u>https://doi.org/10.1007/s10658-012-0024-3</u>
- Indian Horticulture database, 2014. Ministry of Agriculture, Government of India, Institutional Area, Sector-18, Gurgaon, Website : www.nhb.gov.in.
- Indian pharmacopoeia, 1996. Ministry of Health Govt. of India Publication. New Delhi.
- Jones, D. R. 2009. Disease and pest constraints to banana production. *ISHS Acta Horticulturae*, 828: 21–36. https://doi.org/10.17660/ActaHortic.2009.828.1

- Mohapatra, D., Mishra, S. and Sutar, N. 2010. Banana and its by-product utilization: an overview. J. Sci. Ind. Res., 69:323-329.
- Neela, F. A., Sonia, I. A. and Shamsi, S. 2014. Antifungal activity of selected medicinal plant extract on *Fusarium oxysporum* Schlechtthe causal agent of Fusarium wilt disease in Tomato. *American Plant Sciences*, 5: 2665-2671. https://doi.org/10.4236/ajps.2014.518281
- Patil, R.K., Patel, K.D., Sharma, A., and Pathak, V.N. 1995. Inhibitory effect of *Ocimum sanctum* extract on fruit rot fungi. *Indian J. Mycol. Plant Pathol.*, 22(2):190-200.
- Perez, C., Pauli, M. and Bazerque, P. 1990. An antibiotic assay by the well agar method. *Acta Biol. Med. Exper.*, 15: 113 115.
- Ploetz, R. C. 2006. Fusarium wilt of banana is caused by several pathogens referred to as Fusarium oxysporum f. sp. cubense. Phytopathology, 96: 653-656.PMid:18943184 https://doi.org/10.1094/PHYTO-96-0653\_
- Ploetz, R.C. and Churchill, A. C. L. 2011. Fusarium Wilt: the Banana disease that refuses to go away. Proc. Int'l ISHS-ProMusa Symp. On Global Perspectives on Asian Challenges Eds.: I. Van den Bergh, Acta Hort., 897: 519- 526. https://doi.org/10.17660/ActaHortic.2011.897.73.
- Ploetz, R.C., Kema, G. H. and Ma, L. J. 2015. Impact of diseases on export and small holder production of banana. *Annu. Rev. Phytopathol.*, 53:269-88.PMid:26002290 <u>https://doi.org/10.1146/annurev-phyto-080614-120305</u>
- Pradhanang, P.M., Momol, M.T., Olson, S.M. and Jones, J.B. 2003. Effects of plant essential oils on *Ralstonia solanacearum* population density and bacterial wilt incidence in tomato. *Plant Dis.*, 87: 423 - 427. https://doi.org/10.1094/PDIS.2003.87.4.423 PMid:30831840
- Singh, P. and Srivastava, D. 2013. Phytochemical screening and *in vitro* antifungal investigation of *Parthenium Hysterophorus* extracts against A. alternata. Int. Res. J. *Pharm.*, 4: 190-193. https://doi.org/10.7897/2230-8407.04742
- Siva, N., Ganesan, S., Banumathy, N. and Muthuchelian, 2008. Antifungal effect of leaf extract of some medicinal plants against *Fusarium oxysporum* causing wilt disease of *Solanum melogena* L. *Ethnobot. Leaflets*, 12: 156-163.
- Tapwal, A., Nisha, Garg, S., Guatam, N. and Kumar, R. 2011. In vitro antifungal potency of plant extracts against five phytopathogens. Braz. Arch. Biol. Technol., 54(6): 1093-1098. https://doi.org/10.1590/S1516-89132011000600003

P - ISSN 0973 - 9157 E - ISSN 2393 - 9249